

Contents lists available at ScienceDirect

Renewable and Sustainable Energy Reviews

journal homepage: www.elsevier.com/locate/rser



The diverse applications of water hyacinth with main focus on sustainable energy and production for new era: An overview



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ARTICLE INFO

Article history: Received 2 September 2013 Received in revised form 30 June 2014 Accepted 3 September 2014

Keywords:
Biofuel
Biomass
Bioremediation
Briquette
Renewable energy
Water hyacinth

ABSTRACT

Water hyacinth was introduced as an ornamental crop in many countries more than a century ago, due to its attractive appearance and aesthetical value in the environment. Unfortunately, the flowers developed into invasive species due to their adaptability for a wide range of fresh water ecosystems and their interference with human activities. In the 21st century, they were considered as an alternative to fossil fuels, as many researchers found them capable of converting their content into fuel energy at less cost and recognized as an eco-friendly product. As water hyacinth is among the group of fastest growing plants, its biomass has the potential to become a potential renewable energy source and replace conventional fossil fuels, perhaps during the next decade. This is an essential mission to overcome the depletion of energy sources and also to fulfill the increasing demand of world energy. Instead of fuel energy, the dried biomass can also be fabricated as briquettes, which is suitable as co-firing agent in coal power plant. Thus, in future compacted biomass residues produced in the form of briquettes may decrease the dependence of coal to provide more energy The other application of water hyacinth into a co-compost material such as soil amendment to the sandy soil, can improve hydro-physical, chemical parameters of soil and will supply the growing crops with several nutrients. Water hyacinth has also drawn attention due to its bioremediation ability, capable of removing pollutants from domestic and industrial waste water effluents. Thus, the issue of water hyacinth should be evaluated from energy, engineering as well as environmental perspectives. In this review, the potential uses of water hyacinth are being classified and discussed.

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1. Introduction

Water hyacinth (*Eichhornia crassipes*) is a noxious weed that has attracted worldwide attention due to its fast spread and crowded growth, which leads to serious problems in navigation, irrigation, and power generation. It is also renowned as a nonnative, invasive and free-floating aquatic macrophyte. Moreover, water hyacinth due to their abundant and uncontrolled growth in open pond, irrigation and other water bodies, are frequently noted in the literature as one of the world's most problematic weeds [1]. It is a free floating aquatic plant well known for its production abilities and exclusion of pollutants from water. It is able to rapidly grow to very high density of (over 60 kg/m²); this means entire clogging of water bodies can occur, which in turn may have unfavorable effects on the environment, human health and economic development [2–4].

The shoot system covers the surface of water body to capture the sunlight thereby obstructing the entry of sunlight into water, which is required by algae and other organisms present in the water to survive. This leads to a reduction in the growth of algal population and thus disturbs the ecological balance [5]. The mature plant consist of long, pendant roots, rhizomes, stolons, leaves, inflorescences and fruit clusters. The plants are up to 1 m high although; 40 cm is the usual measured height. The inflorescence allows 6-10 lily-like flowers, each one being 4-7 cm in diameter. The stems and leaves contain air-filled tissue which provides the plant its substantial buoyancy. The vegetation reproduction is asexual and takes place at a rapid rate under superior conditions [6]. It can tolerate drought conditions well because it can survive in moist sediments up to several months [7]. At an average the annual productivity of 50 kg/m² dry water hyacinth (ash-free) is in tones per hectare per year [8] and water hyacinth is one of the most productive plants in the world [2]. It can double its size within five days and a mat of medium sized plants may contain two million plants per hectare and can weigh approximately 270-400 t. Also [9] investigated that the growth rate of water hyacinth under favorable conditions can reach up to 17.5 metric tons per hectare per day. These figures also indicate that the plant may interfere with the localized problems, such as navigation, recreation, irrigation, and power generation [10].

The water hyacinth replicates sexually by seeds and vegetatively with the help of budding and stolen production. For rapid spreading, the vegetative promulgation is more important [11]. Daughter plants grow from the stolons and the doubling times have been reported of about 6–18 days. Under favorable conditions of temperature and nutrient availability, the vegetative propagation is very fast. There are seven species of water hyacinth available including: E. azurea - anchored water hyacinth, E. crassipes common water hyacinth, E. diversifolia – variable leaf water hyacinth, E. paniculata - Brazilian water hyacinth. Clonal plants such as E. crassipes might enhance light interception via horizontal growth of stolons or rhizomes and a situation of new ramets, in less shaded microsites [12]. Water hyacinth is successful owing to their life cycle and survival strategy that gives it a competitive edge over other species. Its adaptability at various ecological conditions makes obliteration of this plant virtually impossible [13].

The plant has black, tough roots and when it irregularly becomes stranded in sludge it may appear rooted. Its growth rate is amongst the highest of any plant known, and its population can double in as low as 12 days [14]. A study in Louisiana in 1948, showed that ten plants were able to vegetatively replicate 1610 plants in three months [15] also the growth rate has been calculated in other countries to be an increase in biomass of 400–700 t per hectare per day, or an increase in water area coverage by a factor of 1.012–1.077 per day. Surroundings for water hyacinth have ranged from low temporary ponds, marshes and sluggish flowing waters to large lakes, rivers and reservoirs [1].

The water hyacinth plants can withstand both high acidic and alkaline conditions, but more effervescent growth is supported by neutral water bodies [1]. As assumed by Aquatic Ecosystem Restoration Foundation [16] there is a logistic growth model in the analysis of water hyacinth population dynamics at temperate and tropical zones. Their results revealed that the growth rate in temperate regions differ with seasons. In tropical zones the intrinsic rate of growth for the weed was estimated in the range 0.04–0.08 dry weight per m² per day. It grows over a wide variety of wetland types and prefers nutrient-enriched waters. However, it can tolerate considerable variation in nutrients, temperature and pH levels. The optimum pH for growth of water hyacinth is 6-8. It can grow in wide range of temperatures ranging from 10 to 40 °C (optimum growth at 25-27.5 °C) but it is also thought to be coldsensitive [17]. Salinity is the main obstacle for the growth of water hyacinth in coastal areas [18]. High level of salinity in wastewater can limit the growth of water hyacinth and other aquatic macrophytes [19]. Research performed by De Casabianca et al. [20] showed that water hyacinth would tolerate salinity at less than 10 ppt. In rural areas, water hyacinth could be used as an integrated approach for decentralized wastewater treatment systems coupled to biogas and compost production from the consequential biomass production [8].

Water hyacinth harvests have been put into different valuable uses in several countries. Methods of converting the plant material into valuable products have emerged [19]. This review paper highlights water hyacinth function with the ultimate attention on its utilization for energy and engineering fields conducted in the last three decades. Based on these noteworthy research realizations it is desirable to recognize as an administration strategy to adjust in the commercial activities.

2. Characteristics of water hyacinth

Fresh plant of water hyacinth contains 95.5% moisture, 0.04% nitrogen, 1.0% ash, 0.06% P_2O_5 , 0.20% K_2O and 3.5% organic matter. On zero-moisture basis, it has 75.8% organic matter, 1.5% nitrogen and 24.2% ash. The ash contains 28.7% K_2O , 1.8% Na_2O , 12.8% CaO, 21.0% Cl, and 7.0% P_2O_5 . The crude protein (crude protein = amount of nitrogen \times 6.25) using Kjeldahl method contains, per 100 g, 0.72 g methionine,4.72 g phenylalanine, 4.32 g threonine, 5.34 g lysine, 4.32 g isoleucine, 0.27 g valine, and 7.2 g leucine [21].

3. Application of water hyacinth

Water hyacinth consisting of high percentage of water, fibrous tissue, high energy and protein content can be used for a variety of useful applications. A number of possible uses of the plant includes in the field of bio-fuel production, biomass and energy, waste water treatment, compost and fertilizer, animal feed, furniture, with special focus to the application of water hyacinth for energy sector considering the fuel crisis and the urgency to go for the alternative way of utilizing energy are being discussed extensively in this review paper.

4. Water hyacinth as alternative biomass resource for energy production

Today fossil fuel depletion and more toxic emission formation from combustion of fossil fuel have become the main concerns of energy and environmental societies [22]. The environmental dilemma problems such as climate change, receding of glaciers, increasing the sea level, GHG s effects and lack of biodiversity have been emerged due to lack of appropriate strategy to control the pollutant formation in transportation systems and industrial sectors [23]. Fossil fuels remain as the main source of energy. Recent production of fossil fuels has reached up to 79% compared to other energy resources as shown in (Fig. 1) [24]. However, the demand for fossil fuel as a primary energy source is exceeding its production, due to the rising consumption of fossil fuel energy up to 83% in November 2010.

Due to the population increase energy consumption will also increase in near future (Fig. 2) [24]. Achieving solutions to possible shortage in fossil fuels and environmental problems that the world is facing today require long-term potential actions for sustainable development. In this context, renewable energy resources appear to be one of the most efficient and effective solutions [1]. Bioenergy is now accepted and having the potential to provide

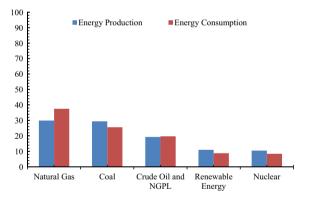


Fig. 1. World primary energy production and consumption in November 2010, per source [24].

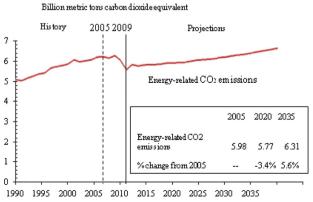


Fig. 2. U.S energy-related carbon dioxide emissions from 1990 to 2035 [24].

a major part of the projected renewable energy provisions required for future [2,3]. As projected in (Fig. 2), the energy related carbon dioxide emission will continue to gradually rise [24].

There are several reasons that drive most of the countries in shifting towards green energy production, which includes wind energy, solar energy, hydropower, biofuel and biomass. Most developed countries will focus on renewable energy that is more adaptable for them, for economical purposes, depending on the geographical location, climate condition and availability of renewable sources. For example, insolation (solar radiation) in the Mojave Desert near Barstow in California is good enough for photovoltaic activity and therefore suitable for solar power plants application [24]. In future, advanced biofuel produced from lignocellulosic biomass can replace biofuel produced from agricultural feedstock [25].

As aquatic plants do not compete with land resources used in arable food crop cultivation and thus are an incentive factor when it comes to biofuel production [26]. However, there are no exact figures available for bio-alcohol production from water hyacinth [27]. The energy crisis of 1970s renewed interest in alcohol production for fuels and chemicals. Ethanol is used in vehicles either as a sole fuel or blended with gasoline and with the growing energy crisis supplemented by environmental concerns, biomethanation of water hyacinth can serve as a biomass-to-energy generation alternative. Water hyacinth management problems and environmental concerns as well as the on-going successful shifting from non-conventional to renewable energy technologies has given an impulse to focus on biogas production.

At present, much focus is on the development of methods to produce ethanol from biomass that possesses high cellulose content. This cellulosic ethanol could be produced from abundant low-value material, including wood chips, grasses, crop residues, and municipal waste. In the recent days, the cellulosic substrates searching has gained a new speed and is continuing, some of it includes water hyacinth, sunflower stalks, etc., which are being explored for ethanol production possibility in different laboratories [28].

The world ethanol fuel production for different continents, 2006–2011 [29] is shown in Table 1 and can be seen that America is dominant in the production of alcohol.

Generally, biofuel represents all of gaseous and liquid fuels mainly extracted from biomass. For example biodiesel, biomethanol, bioethanol and biohydrogen are extracted from biofuel [30]. The privileges of biodiesel not only have convinced governments to take biodiesel in account as an energy resource but also have pursued them to take new strategies to expand biodiesel production. Statistics illustrate that biodiesel production has risen drastically in recent years. Fig. 3 depicts world annual ethanol and biodiesel production from 2005 to 2020 respectively according to OECD and FAO Secretariats [31]. Also biodiesel, bio methanol, bioethanol and biohydrogen as main biomass products have been applied vastly in energy generation. Huge amount of biofuel resources, fluctuation in fossil fuel prices and environmentally friendly characteristics of biofuel combustion process have convinced the governments of tropical countries to invest in the development of biofuel industry [32,33]. It has been proven that the application of biodiesel blends mitigates the rate of CO₂ formation in industrial boilers dramatically [34].

For the development of alternative energy sources, the processing of agricultural wastes into biomass is an appropriate strategy. Water hyacinth is an aquatic plant which can be used as an alternative energy source to wood or other related briquettes. This aquatic plant was harvested, dried and ground into powder before being mixed with starch binder in manually operated briquetting machine. The briquettes produced were sundried and comparative cooking tests were carried out by burning the

Table 1 World ethanol fuel production (million liters). . Source: F.O. Licht (2011)

Region Year						
	2006	2007	2008	2009	2010	2011
Americas	35,625	45,467	60,393	66,368	77,800	79,005
Asia/Pacific	1940	2142	2743	2888	3.183	4.077
Europe	1627	1.882	2.814	3.683	4.615	5.467
Africa	0	49	72	108	165	170
World	39,132	49.540	66.072	73.047	85.763	88.719

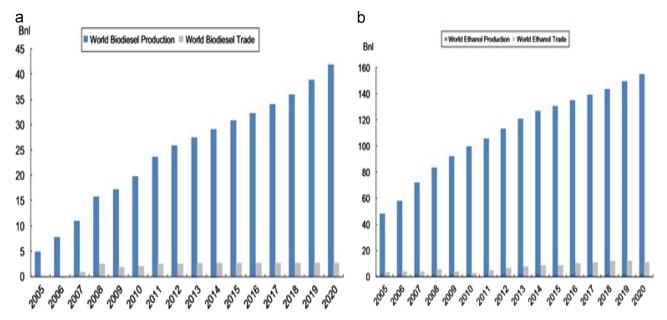


Fig. 3. World biofuel prediction (a) biodiesel and (b) ethanol [34].

briquette in a mold [35]. Forhadlbne et al. [36] have shown the potential of biogas production using cow dung, poultry waste and water hyacinth through anaerobic digestion. These wastes are always available in our environment and can be used as a source of fuel if managed properly. Biogas technology can be a viable development option for developing countries for energy production and substitution if properly managed and marketed.

4.1. Water hyacinth as co-fuel

Many researchers and scientists have attempted different kinds of fuels namely compressed natural gas (CNG), liquefied petroleum gas (LPG), hydrogen, and alcohols. The vegetable oils and alcohols (methanol and ethanol) are favorable renewable liquid fuels. Because of low cetane number, alcohols are not qualified for diesel engines. The poor volatility and low octane number make vegetable oils incompatible for spark ignition (petrol) engines. One possible solution to solve this problem is the use of bio-diesel. Two fuels which can be used as sole fuel or as mixture along with diesel fuel are: straight vegetable oil (SVO) and bio-diesel (which are esters of SVO) [37–41].

Water hyacinth satisfy all the criteria for bioenergy production – it is permanent, plenty available, non-crop plant, biodegradable and has high cellulose content; however its strong disadvantage is that it has over 90% water content which complicates the process of harvesting and processing. The biomass can be subjected towards biogas production as to generate energy for household use in rural areas [42].Water hyacinth is low in lignin content (10%), contains high amount of cellulose (20%) and hemicellulose

(33%) [43-45]. Since a few decades, researchers have found its capability in fermentation process to generate biofuel and bioethanol. A typical biomass from land plants can have 30-50% cellulose, 20-40% hemicellulose and 15-30% lignin, which could hinder its capability in fermentation processes by selected commercial yeast and enzymes. In plants, lignin (composed of phenyl propanoid groups) acts as a polymer around the hemicellulose micro fibrils, binding the cellulose molecules together and protecting them against chemical degradation. Therefore, lignin compounds cannot be converted into sugars and may limit the capability microbiological activity during fermentation process. Therefore, some plants that contain higher lignin content are not useful for biofuel production. Their degradation involves high energy process. As water hyacinth has low lignin, the cellulose and hemicellulose are more easily converted to fermentable sugar therefore resulting in enormous amount of functional biomass for biofuel industry. As suggested by Poddar et al. [46] a new method of extracting ethanol from fermented sugar can be derived by saccharification technique, pretreatment with diluted sulfuric acid and further accelerate the fermentation process by using selected yeast extraction. In certain countries, water hyacinth is successfully utilized for biogas production at different fermentation capacities [47-49]. It is concluded that a mixture of 25% cow dung and 75% of dry water hyacinth could yield the best rate of methane production and specifies the huge potential of water hyacinth as non-conventional energy source with the anticipation that oneton of dry water hyacinth could yield 370,000 L of biogas.

Considering the cost of various feed stock, to reduce the cost of fuel alcohol production, the use of water-hyacinth offers an

opportunity which is freely available in large amount as a substrate. The mixture of cow dung and water hyacinth slurry has proven to produce more biogas than when used alone [50]. The application of fuel ethanol (bioethanol) from biomass involves the step of pre-hydrolysis, hydrolysis, fermentation, and distillation. The hydrolysate resulting after pre-hydrolysis and hydrolysis include changeable amount of monosaccharide's, both pentose's and hexose's, and also a wide range of substances either obtained from the raw material or resulting as the reaction product from sugar and lignin degradation. Many of these compounds may have inhibitory consequence on microorganisms in subsequent fermentation steps [51–56]. The fermentation organism must be capable of fermenting all monosaccharide's present and in addition. survive potential inhibitors in the hydrolysate. The most normally used ethanol producer, Saccharomyces cerevisiae, cannot ferment pentose's, which may represent up to 40% of the raw material. Among the xylose fermenting yeasts Pichia stipitis, has shown promise for industrial application; because it ferments xylose quickly with a high ethanol yield and apparently produces no xylitol [57], and is able to ferment a wider range of sugars (including cellobiose) than Candida shehatae [58]. Also it has been demonstrated that the hydrolysate of water hyacinth biomass (WHB) can be fermented to ethanol using the common baker's yeast Saccharomyces cerevisiae albeit with lesser efficiency. The use of more suitable organisms for fermentation can improve the yield of ethanol. Almost all of the technologies that are employed in the production of bio-ethanol from WHB is less technically intensive and can be operated by non-skilled workers making the strategy suitable for small scale distributed production of fuel ethanol [59].

As shown in Table 2, recent studies have been carried out to optimize the operating parameters as well as the design and method of appropriate technology to convert water hyacinth into biofuel

Many studies were conducted by pre-treatment processes, including acidification technique and numerous methods were developed for this purpose, including physical, chemical or biological processes. However, green ethanol is the preferred, ultimate raw alternative for energy and the best way is to apply biological processes for pretreatment and fermentation. The main issue in biological processes is their time consumption and in order to improve the biological limitation, researchers are currently modifying

biochemistry and bioprocess activities for the fermentation process. As biological processes are eco-friendly and lead to energy savings, one must carefully consider biological processes to preserve the environment and protect the planet which is increasingly under threat [4].

4.2. Water hyacinth as briquette biomass (power plant energy)

Briquetting, also known as biomass densification of agro residues has been practiced in several countries for many years [68]. In the briquetting process, biomass residues will turn into uniform and solid fuel as briquettes which will enhance its properties with more added values. This biomass briquette has the characteristic of higher density and energy content besides having less moisture content compared to their raw materials [69]. Among the commercial biomass briquettes are sawdust briquettes, wood residues and rice husk briquettes which are available in the local market [68,69]. In Kenya, the purpose of briquetting is to deal with the massive growth of water hyacinth and is expected to benefit the lake side communities. As water hyacinth is the fastest growing plant, thus its biomass has the potential to become the renewable energy source in replacing conventional fossil fuel [70].

Biomass waste material is usually very bulky and has a very low density which makes it very difficult to be used in many types of burners and transportation is also very uneconomical [71]. Densification of biomass is one of the solutions to this problem. Briquette quality is evaluated mainly by the briquette density [72]. If produced at low cost and made conveniently accessible to consumers, briquettes could serve as complements to firewood and charcoal and kerosene for domestic cooking and agroindustrial operations, thereby reducing the high demand for them. Besides, briquettes have been noted to have advantages over fuel wood in terms of greater heat intensity, cleanliness, convenience in use, and relatively smaller space requirement for storage [73]. A number of locally available materials have been found to be suitable for briquetting into fuel energy production. These include sawdust, cowpea chaffs, corn cobs, and water hyacinth [74].

Wide development of industrial sectors often results in creating a polluted environment. The availability of coal leads to increased use of fossil fuels worldwide. Easy access to electricity is the basis of higher living standard. As in United States, coal-fired

Table 2Biogas production from various studies using water hyacinth.

Reference	Summary/methods of application	Results/findings
[60]	Pretreatment with fungi or chemicals to increase the biodegradability of water hyacinth.	Untreated water hyacinth: $0.10l g - 1 dm$; <i>Phanerochaete chrysosporium</i> pretreatment: $0.24l g - 1 dm$; pyrogaliol red + <i>P. chrysosporium</i> : $0.31l g - 1 dm$.
[61]	Water hyacinth+night soil (1:3 and 3:1).	Increased biogas production, rich in CH ₄ .
[62]	Investigation of, both experimentally and theoretically using mathematical tools, a fermentative system utilizing water-hyacinth hemicellulose acid hydrolysate as a substrate for ethanol production using <i>Pichia stipitis</i> .	Xylose (72.83%) converted to ethanol with a yield of 0.425 gp/gs and productivity of 0.176 gp/L/h.
[59]	Suitability of feedstock for production of fermentable sugars using cellulase produced on site and testing of acid and alkali pretreatment methods.	Enzyme blends resulted in improvement of saccharification from 57% to 71% with Saccharomyces cerevisiae.
[63]	The mild acid pretreatment and combination of biological pretreatment by white rot fungus <i>Echinodontium taxodii</i> or a brown rot fungus <i>Antrodia sp.</i> 5898 with mild acid pretreatment were evaluated.	Ethanol yield from co-treated water hyacinth achieved 0.192 g/g of dry matter, which increased 1.34-fold than that from acid-treated water hyacinth (0.146 g/g of dry matter).
[64]	Two-sequential steps of acid hydrolysis (10% sulfuric acid) and yeast (<i>Candida shehatae</i> ; xylose-fermenting yeast) used for bioconversion of water hyacinth to liquid ethanol.	
[65]	The acid pretreatment and enzymatic hydrolysis was used to ferment sugar to ethanol. Hydrolysis and fermentation (SHF) studies were carried out separately to produce ethanol from water hyacinth leaves.	Ethanol concentration of 3.39 (g/l), percentage of theoretical ethanol yield of 96.07%, ethanol yield of 0.25 g/g and the volumetric productivity of 0.221 g/l h obtained.
[66]	The effect of physical (subcritical water) and chemical (acid and alkali) pretreatment on conversion of lignocellulose (cellulose, hemicellulose) in water hyacinth (WH) was investigated.	Combination of acid or alkali pretreatment with enzyme treatment resulted in drastic increase of sugar in samples (up to 31.2 and 22.9%) w/w, after fermentation, up to 60% of sugar in the sample converted to ethanol.
[67]	The optimization of pretreatment process was conducted for the enzymatic hydrolysis of lignocellulosic biomass (water hyacinth).	The order of catalytic effectiveness for hydrolysis yield was found to be phosphoric acid > maleic acid > sulfuric acid.

Table 3 Briquette production from water hyacinth.

Reference	Summary/methods of application	Results/findings
[35]	Comparison of cooking time obtained were made along with the time obtained, when same quantities of food were cooked using a conventional kerosene stove.	Use of water hyacinth briquette took longer time for cooking purpose, when compared to kerosene stove. Water hyacinth biomass can serve as an alternative energy source.
[91]	Assessment of electric power generation using water hyacinth (WH) and agricultural waste.	Produced electricity ranging from 1.38 to 1.41 kWh/m³. 1 t of WH can produce average 13.3 m³ of biogas and 18.35–18.75 kWh electricity
[86]	Effect of compaction pressure, binder proportion, particle size on ignition time and burning rate of fuel briquettes produced were carried out with the mixture of water hyacinth and plantain peel.	Compaction pressure and binder proportion caused decrease in burning rate.
[92]	Quality of fuel briquettes made from sewage sludge mixed with water hyacinth was investigated.	Highest calorific value of (3362.9 cal/g) was obtained at 1:3 ratio and provided highest compressive strength value of (4545 N).
[89]	Assess the combustion characteristic of briquettes produced from mixed water hyacinth and plantain peels as binder, mangrove wood, charcoal and <i>Anthronotha macrophylla</i> (firewood).	The results confirm the possibility of utilizing water hyacinth as fuel briquettes as good source that can support combustion, to provide high material strength as well as high value of combustible fuel.
[93]	Use of water hyacinth briquettes as alternative to the locally available wood fuel.	Water hyacinth briquette has greater amount of moisture content, similar amount of volatile matter, and much greater ash content, significantly much less fixed carbon and much lower calorific value when compared to the local wood fuel.

power plants generate half of the electricity [75]. This fact greatly contributes towards the environmental problem related to the emissions of SO₂, NOx and greenhouse gas (CO₂) from the activity of co-firing coal. Thus, co-firing biomass with coal in the existing coal fired boilers will reduce the emission of greenhouse gases [76]. In their research it was found that the co-firing woody biomass and bituminous coal with the proportion of 60:40 showed great reduction of SO₂, NOx and suspended particulate matter (SPM) as compared to co-firing bituminous coal alone. Unfavorable emissions can be reduced up to 15–20% by partial combustion of biomass and coal in the existing pulverized coal-fired boilers [77]. Biomass co-firing can reduce the share of emissions per unit energy produced [77] with small capital investment as compared to single coal firing [78].

The best selection of raw material for briquetting is based on their moisture content, ash content, flow characteristic and particle size. Most of biomass residues possess lower ash content, but at the same time they have higher potassium content which tends to devolatilize during combustion and become condensed on superheated surfaces [68,79]. Thus, it does not mean that biomass with lower ash content will not show any slagging behavior even though in general, biomass with greater ash content would form great slagging behavior [68]. In one study, [80] it is found that there was no significant difference in terms of calorific value between charcoal briquette from water hyacinth and mixture of charcoal including water hyacinth powder. Many researchers have reported on the combustion properties of briquettes fuel for various agricultural waste products such as charcoal briquettes from neem wood residue [81]. Also, production of fuel briquettes from waste paper and coconut husk [82], rice husk and saw dust [83], composite sawdust briquettes [84], palm kernel briquettes [85] and mixture of water hyacinth and plantain peel [86].

As stated by [87], biomass briquetting can enhance the agro residues characteristic for feeding into furnaces and combustion. While [88] suggested that less compact form of agro residues which are transforming into briquettes form is technologically more efficient when compared to direct firing. It is also reported by [88], that 67% of the total energy consumption in India consists of coal. Besides, higher production of ash (low calorific value), a need to low cost transportation and bad environmental impact were the main reasons for India to substitute coal into briquetting, also because of its abundant agricultural residues. Rotimi et al. [89] demonstrated the use of briquettes produced from water hyacinth to be considered as environmental friendly and to reduce health hazard associated with the use of fuel—wood and charcoal.

Furthermore, when a piece of wood was mixed into briquette it showed greater expansion in cooking time. This suggests that water hyacinth biomass in the form of briquette can be used as charcoal substitute for industrial purposes. Among the third world countries, India is the only country that has successfully developed their briquette sector [68]. As discussed by Emerhi [90], it was found that a charcoal and sawdust based briquette can produce a hot, smokeless and longer time of fire. Table 3 shows some recent studies of using water hyacinth as fuel briquette, charcoal substitute and electric power generation.

5. Other applications

5.1. Wastewater treatment (phytoremediation)

Harvesting aquatic plants to withdraw nutrients from waste water is one of the early methods to reduce pollution in lakes. It is pointed out that all aquatic plants can serve this purpose. However, small plants, like phytoplankton, or submerged plants are more difficult and expensive to harvest than the floating and emergent vascular plants [94]. Water hyacinth has the ability to clean up various contaminated waters [95-97]. In the past decades, the application of Waste Stabilization Pond (WSP) has attracted attention to treat swine effluent. Both laboratory and field studies shows that water hyacinth is able to reduce a variety of pollutants present in the swine wastewater [98]. Numerous researchers [99-101], have tested Eichhornia crassipes, Pistia stratiotes, Salvinia rotundifolia and Lemna minor and found that water hyacinth (E. crassipes) has greater capacity of nitrogen and phosphorus removal. Phosphorus pollution in aquatic environment is generally recognized to be from three major sources: industry, agriculture and domestic sewage [102]. E. crassipes has been known to support its growth contaminated water because of its settlement action and absorption capacity. The high productivity rate of water hyacinth is one of the most important reasons why it has been universally used in Southern France for the treatment of industrial waste waters [103]. As mentioned by Williams [104] the efficiency of nitrogen removal rate is from 10 to 90%. Also research over the past few decades has demonstrated that some floating plants, such as water hyacinth (E. crassipes), water lettuce (Pistia stratiotes), pennywort (Hydrocotyleum bellata), duckweed (Lemna minor), water peanut (Alternantheraphiloxeroides) and lidded cleistocalyx (Cleistocalyx operculatus), have the greatest effects on purifying eutrophic water [105-107]. Therefore, water hyacinth and duckweed were tested in the treatment of pig and dairy manure-based wastewater [15,108].

5.1.1. Removal of organic pollutants

Among the floating aquatic plants, water hyacinth has been extensively studied at laboratory, pilot and larger scale for removing the organic matter present in wastewater [109]. Even though water hyacinth is a persistent plant in most of the countries all over the world; it is also being used as a resource in agricultural and waste management process [20]. As estimated by Wu et al. [110], the practical use of water hyacinth grown in natural water channels or ponds is mainly for water purification. They suggested shorter harvesting intermission to obtain more yield of the biomass. As indicated by [111] in the treatment of dairy wastewater, water hyacinth was discovered to be more effective. The wastewater was treated with water hyacinth for a period of 25 days and various physio-chemical parameters were analyzed according to the series of treatment. After 25 days, total solids, calcium, magnesium and total hardness were reduced by 37, 5, 47.5, 54 and 33%, respectively. Water chloride, chromium, nitrous nitrogen, nitric nitrogen, pH, alkalinity, COD, BOD, bicarbonates were also reduced considerably. This phytoremediation technology is suitable in treating industrial wastewater since it can be used to treat contaminated soils, groundwater and wastewater in both low cost application and technology [112].

As reported by Jayaweera et al. [113], water hyacinth growth under the different nutrient conditions for Fe enriched wastewaters in batch-type constructed wetlands have been studied base on phytoremediation efficiencies. At the 6th week with a highest accumulation of Fe, 67 mg/kg dry weights, the obtained results showed the highest phytoremediation efficiency of 47% during optimum growth. Table 4 shows the some recent studies about application of water hyacinth for treatment of any type of waste water.

5.1.2. Removal of toxic pollutants and heavy metals

With global heavy metal contamination on the rise, plants that can process heavy metals might provide efficient and ecologically sound approach to sequestration and removal [119–121]. The phytoremediation of metals is a cost-effective green technology based on the use of metal-accumulating plants to remove toxic metals,

including radionuclide, from soil and water. Phytoremediation takes advantage of the fact that a living plant can be considered as a solar-driven pump, which can extract and concentrate particular elements from the environment [115]. The root of the plant will be absorbing the metal pollutant that restrain in the wastewater and enhance the quality of water [15]. Water hyacinth has drawn attention as a plant capable of eliminating pollutants, including toxic metals from surface water. Reduction of heavy metals in situ by plants may be a functional detoxification mechanism for phytoremediation. Numerous studies were conducted to determine the phytotoxic effects and uptake capacity of heavy metals by water hyacinth [64,122–124].

Aquatic macrophytes have frequently been used to monitor freshwater pollution by heavy metals and pesticides. In tropical and subtropical regions, because of its abundance and the large biomass produced, the water hyacinth (*Eichhornia crassipes*) has been studied especially for this purpose. The ecology and the practical use of this species were comprehensively revised by Delgado et al. and Klumpp et al. [125,126].

The root structures of water hyacinth (and other aquatic plants) supply an appropriate environment for aerobic bacteria to function in sewage systems. Aerobic bacteria feed on nutrients and produce inorganic compounds which in turn provide food for the plants. To prepare rich and valuable compost, the plants grow rapidly and can be harvested. Water hyacinth has also been used for the removal or reduction of nutrients, heavy metals, organic compounds and pathogens from water [1].

Water hyacinth is not only capable to absorb and accumulate heavy metals, but also it can tolerate toxicity by converting it from chemically-active toxic status to inactive and nontoxic status. For example, [90] estimated the removal of chromium (III) from aqueous solution by water hyacinth. The results proved chromium removal rate at 10 mg Cr/1 solution, which the recovery was monitored at 87.52%. However in higher concentrations (e.g. 50 or 100 mg/1), the water hyacinth response in a weak performance that concluded higher ratios of water hyacinth mass/solution volume lead to higher solution decontamination. The absorption of Cr (III) by water hyacinth was lowered by decreasing the pH solution. The Cr (III) removal by water hyacinth was not influenced by the temperature in the range 17.5–26.0 °C or by the dissolved oxygen concentration in the solution between 4.7 and 6.8 mg/l.

In a study in Dhaka, Bangladesh, Water hyacinth plants from a pond were dried in air and a fine powder was prepared from the

Table 4Phytoremediation of nutritionally rich wastewater by water hyacinth.

Reference	Wastewater source	Main pollutants	Results	Comments
[109]	Piggery wastes (tertiary treatment)	COD, BOD, TN, TP	50% reduction in all parameters at 110 kg TN/ha/d and 20 d HRT	Use of harvested WH as animal diet
[114]	Sewage treatment	Metals, BOD, nitrates and phosphates	Removal of metals (20–100%) BOD (97%); nitrates and phosphates (> 90%)	Combination of WH, duckweed and blue-green algae
[115]	Dairy effluent	Nitrogen and phosphorus	Removal of N (72%) and P (63%)	Combination of WH and Duck weed gave N (79%) and P (69%) removal
[15]	An aerobically digested flushed dairy manure wastewater (1:1 dilution)	Total nitrogen (TN), total phosphorus (TP), NH4 +, EC, Na+	Reduction (%) of TN (91.7), NH4+(99.6), TP (98.5) coupled with reduction in EC and Na+ $$	WH gave best performance While poly culture (of 3 spp.) ranked 2nd
[116]	Duck farm	COD,TP. TN,	COD removal 64.44 (%) TP removal 23.02 (%) TN removal 21.78 (%)	Remarkable removal from duck farm
[117]	Swine wastewater	Nitrogen and phosphorus	The adsorption efficiency was about 36% upon saturation	Showed much greater NH ₃ –N reduction efficiency
[118]	Fish farm wastewater	pH, turbidity, (DO), (COD), (BOD), nitrite phosphate (PO $_4$ ³⁻ , nitrate (NO 3 -), nitrite (NO 2 -), ammonia (NH $_3$), and total kjedahl nitrogen (TKN)	pH ranging from 5.52 to 5.59 and from 4.45 to 5.5, reduction of turbidity were 85.26% and 87.05%, similar reductions were observed in COD, TKN, $\rm NO^{3-}$, $\rm NH_3$, and $\rm PO_4^{3-}$	Removal of aquatic macrophytes from water bodies is recommended for efficient water purification.

Table 5Uptake of some heavy metals by water hyacinth.

References	Type of heavy metal removal	Findings	Comments
[131]	Arsenic, chromium, mercury, nickel, lead, zinc	Six different concentrations ranging from 5 mg/l to 50 mg/l were studied. It was observed that in aqueous solutions containing 5 mg/l of arsenic, chromium and mercury the maximum uptake were 26 mg/kg, 108 mg/kg and 327 mg/kg of dry weight of water hyacinth respectively.	At lower concentrations(5 mg/l) of heavy metals, the plant growth was normal and removal efficiency was greater.
[132]	Cadmium (Cd), lead (Pb), copper (Cu), zinc (Zn), and nickel (Ni)	The concentrations in the root tissue were found in the order of $Cu > Zn > Ni > Pb > Cd$. The absorption capacity for water hyacinth was estimated at 0.24 kg/ha for Cd, 5.42 kg/ha for Pb, 21.6 kg/ha for Cu, 26.2 kg/ha for Zn, and 13.5 kg/ha for Ni.	Translocation ability was defined as the quantity of Cu, Pb, Cd, Ni, and Zn translocated in the plant's tissues and expressed as a root/shoot ratio.
[133]	Cadmium (Cd) and zinc (Zn)	Highest concentration of metals in roots (2040 mg/kg for Cd and 9650 mg/kg for Zn). However, relatively little Cd (113 mg/kg) was translocated to the shoot, while Zn was translocated at a much higher concentration (1930 mg/kg).	
[134]	Chromium,	Water hyacinth has tremendous potential to absorb heavy metals from the textile wastewater as it resulted in 95% reduction of chromium, 97% in zinc and 94% reduction in copper.	ANOVA analysis showed a significant (p < 0.05) reduction in pollutants with the passage of time, especially in textile industry wastewater.
[128]	Arsenic	The amount of arsenic remaining in solution was found to be less than	Removal of arsenic solution concentration of 1500 mg l_1, approximately 30 g of dried roots were required to remove 1500 mg of arsenic from 1 l of water in 24 h.
[135]	(Cd2+), (Cu2+), (Pb2+), (Zn2+)	10 kg ⁻¹ which is the WHO guideline limit value for As in drinking water. The equilibrium data were correlated with Langmuir and Freundlich Isotherm models. Based on the Langmuir model the maximum adsorption capacities were found to be 3.2. 18 16 and 13 mg/g' for cadmium, copper, lead and zinc respectively.	Maximum removal of metal ions took place at pH 4–6.

roots. From a solution containing 200 g of arsenic per L within 60 min of exposure to the powder, more than 93% of arsenite and 95% of arsenate were removed. The arsenic concentration remaining was less than the WHO drinking water guideline value of 10 g/L. Earlier Misbahuddin and Fariduddin [127], had noted that water hyacinths removed arsenic when placed in arsenic contaminated water for 3–6 h. The extent of arsenic removal depended on the arsenic concentration present, the amount of water hyacinth used, the exposure time and the presence of air and sunlight. Also Shaban et al. [128] reported that whole plants were more effective in removal of arsenic compared to fibrous roots alone.

Also water hyacinths (E. crassipes) were used as a pollution monitor for the simultaneous accumulation of arsenic, cadmium, lead and mercury [129]. The plants were harvested and rinsed with tap water after 2 days of cultivation in tanks containing 10 ppm each of As, Cd, Pb and Hg in aqueous solution. Also for each type of metals, the leaves and stems were separated and analyzed. The ratio of the arsenic and mercury concentrations in the leaves to that of the concentrations in the stems was found to be 2:1. Cadmium and lead showed a concentration ratio of about 1:1 in the leaves versus the stems. At 0.3 mg/g of dried plant material, the arsenic concentration in leaves was the lowest of all the metals. The leaf concentration of cadmium was highest at 0.5 mg/g of dried plant material. Arsenic removal by water hyacinths (E. crassipes) was also reported by Low and Lee [130]. Table 5 shows some recent studies about uptake of some heavy metals by water hyacinth for treatment of any type of waste water.

6. Composting and fertilizer

Water hyacinth can be used in farming as an organic fertilizer and as a mulch crop. There has been an increase in demand for organic foods, particularly in the developed world. The plant also contributes to protect soil moisture and nutrient recuperation. The plants can be turned into compost and used as a fertilizer. The

plant tends to retain most of the nutrients at a dry condition [136]. The time taken in composting is only 30 days compared to other crop plants, which can take up to 2–3 months [137].

The water hyacinth can be used on the land either as surface mulch or as compost. Mulching field crops with water hyacinth was found to increase the production of lady finger (67%), potato (14%) and tomato (90%) as compared to control (no mulching) treatment [138,139]. However Lenzi et al. [140] observed that mulching with water hyacinth when treated with maximum rates of glyphosate, and 2,4-D herbicides reduced growth of tomato as compared to untreated water hyacinth when used as mulch. Therefore, while using water hyacinth for mulching or composting purpose more intensive care is required such as not to spray with herbicides. Predictable composting, which is suitable for labor intensive, low capital production can be done by mixing dried plant with ash, soil and some animal manure/organic municipal waste. Vermicomposting of water hyacinth is more valuable because the water hyacinth losses its capability to reproduce vegetatively after it has passed through the earthworm gut [141].

The biomass of water hyacinth can be used directly as green fertilizer as compost. Furthermore the assimilated vegetative waste from biogas generation as specified above can also be collected for using it directly on the farm. Otherwise, these materials could be mixed with other organic materials before use. The crude powder obtained from the root of water hyacinth has successfully been used to support crop production in economic crops such as vegetables [142]. As mentioned by Oguniade et al. [143] water hyacinth can be rich in nitrogen, up to 3.2% of DM and have a C/N ratio around 15. Also Water hyacinth, due to its abundant growth and high concentrations of nutrients, has a great potential as fertilizer for the nutrient deficient soil.

7. Animal feed (livestock)

The lack of animal protein with increasing cost of food production coupled with rapid population growth demanded the search for non-conventional sources of protein such as leaf protein concentrate (LPC) from water hyacinth [144]. This plant in arrangement with concentrate of other feeds has established to be a good quality protein source for animal feed [145]. The high water and mineral content of water hyacinth indicates that the nutrients in water hyacinth are appropriate to some animals as feed [142]. Boiled and chopped water hyacinth along with vegetable waste, rice bran, copra cake and salt is used to make suitable feed for pigs in China. When the plant is sun-dried, it has been found to be rich in protein, vitamins and minerals and serves as a high quality feedstock for some non-ruminant animals, poultry and fishery in Indonesia, China, the Philippines and Thailand [146,147].

In Malaysia, Indonesia, Philippines and Thailand the water hyacinth is used as feed for pigs, ducks and fish [148]. As indicated by Oguniade et al. [143] because its dry matter has high crude protein (18%) and low acid detergent fiber (33%) contents the water hyacinth has potential as a roughage source for ruminants. It is also reported that when grass crop (Ctenopharyngodon idella) were fed diets containing from 0 to 100% water hyacinth meal, weight gain and protein efficiency ratio decreased as the amount of water hyacinth meal increased. Water hyacinth has also been used indirectly to feed fish. Dehydrated water hyacinth has been added to the diet of channel catfish fingerlings to increase their growth [149]. It has also been noted that decay of water hyacinth after chemical control releases nutrients which promote the growth of phytoplankton with subsequent increase in fish yield [127]. The effects on egg quality with water hyacinth as duck feed (cattle feedstock), eggshell thickness and intensity were evaluated based on statistical analysis [116]. As well as encouraging digestion and absorption in the ducks, water hyacinth is rich in protein and minerals, especially calcium, which reached 2.0% of dry matter, and thus it could increase the eggshell strength. Because the experimental and control groups had the same eggshell relative weights, this specified that adding water hyacinth to the diet increased egg weight and consequently increased the eggshell weight [150].

8. Furniture

Water hyacinth can be used for furniture-making. So far it has not been figured out by any other researchers for academic matters even though commercial products have been developed in India, Thailand, China and Indonesia. Since no other researcher utilizes their potential previously, the furniture-making from water hyacinth is a novelty research. Using water hyacinth for furniture making is still difficult due to the higher material quality demand and difficulty of process making. However, as discussed by Jafari [19], biomass can be turned into durable, esthetic furniture and handicrafts professionally. In the Philippines, this weed is dried and the stalks are weaved into baskets and mats. Also, in India, similar domestic goods are manufactured from this weed such innovative decorative articles are known among visitors.

9. Conclusion

By referring to the latest research on the water hyacinth has promoted our awareness about the basic as well as applied characteristics of this plant. This information will help to develop any future utilization of water hyacinth. A suitable large-scale utilization could supply as a positive advance to control the spreading of the water hyacinth. The paper has discussed the adaptability of the water hyacinth to deal with domestic/industrial

wastewaters of various origin and nature and shown the efficiency of this plant for removal of pollutants from wastewater. The water hyacinth can also serve as an alternative source of energy.

According to literature, the most and beneficial usage of water hyacinth is biofuel production, mainly in Brazil, India and some African countries. It is found that 1 kg of cellulose yields 1.1 kg of glucose and 1 kg of cellulose yield 0.56 kg of ethanol.

Other consumption options including water hyacinth-based power plant energy, compost/fertilizer and animal feed production also make the engineering feature important in many commercial ways. The focus on future outlook should be power plant energy. It can help to decrease the use of fossil fuels. The use of water hyacinth in developing countries for animal feed may help solve some of the nutritional limitations in these countries. Treating the presence of abundant water hyacinth and using its significant heat content as an ideal opportunity to sensitize local communities for serving it as alternative biomass with economic and environmental advantages will be an added value.

The new idea can be to use the plant for several purposes at the same time. For example, after water hyacinth phyto-remediation and uptake nutrient from waste water, the biomass can be used in power plant energy or biofuel production. However, until now the utilization has not developed into large scale activities other than localized cottage industries or to support poor communities in subsistence continuations such as the production of biogas. Government could further break the barrier and provide means to educate people about usage of this non-fossil source of energy and also support the scientific research to further explore the plant potential.

References

- [1] Gopal B. Aquatic Plant Studies 1. Water Hyacinth. Oxford; Elsevier; 1987; 471.
- [2] Fernández OA, Sutton DL, Lallana VH, Sabbatini MR, Irigoyan JH. Aquatic weed problems and management in South and Central America. In: Charudattan R, editor. Aquatic weeds the ecology and management of nuisance aquatic vegetation. New York: Oxford University Press; 1990. p. 406–25.
- [3] Epstein P. Weeds bring disease to the east African waterways. Lancet 1998:351:577.
- [4] Ganguly A, Chatterjee PK, Dey A. Studies on ethanol production from water hyacinth a review. Renew Sustain Energy Rev 2012;16:966–72.
- [5] Tiwari S, Dixit S, Verma N. An effective means of biofiltration of heavy metal contaminated water bodies using aquatic weed *Eichhornia crassipes*. Environ Monitor Assess 2007:129:253–6.
- [6] Matai S, Bagchi DK. Water hyacinth: a plant with prolific bioproductivity and photosynthesis. In: Gnanam A, Krishnaswamy S, Kahn JS, editors. Proceedings of the Proceedings of the International Symposium on Biological Applications of Solar Energy, India: Macmillan: 1980. p. 144–8.
- [7] Center TD, Hill MP, Cordo H, Julien MH. Water hyacinth. In: Van Driesche R, et al., editors. Biological control of invasive plants in the Eastern United States, 4. Washington, DC: USDA Forest Service Publication FHTET; 2002. p. 41–64.
- [8] Abbasi SA, Ramasamy EV. Biotechnological methods of pollution control. Hyderabad: Orient Longman (Universities press India Ltd.); 1999; 168.
- [9] Kunatsa T, Madiye L, Chikuku T, Shonhiwa C, Musademba D. Feasibility study of biogas production from water hyacintha case of lake Chivero – Harare, Zimbabwe. Int J Eng Technol 2013;3(2):119–28.
- [10] Cohen Y, Plaut Z, Meiri A, Hadas A. Deficit irrigation of cotton for increasinggroundwater use in clay soils. Agron J 1995;87:808–14.
- [11] Wilson JR, Holst N, Rees M. Determinants and patterns of population growth in water hyacinth. Aquat Bot 2005;81:51–67.
- [12] Verma R, Singh SP, Ganesha Raj K. Assessment of changes in water hyacinth coverage of water bodies in northern part of Bangalore city using temporal remote sensing data. Curr Sci 2003:795–804.
- [13] Methy M, Alpert P, Roy J. Effects of light quality and quantity on growth of the clonal plant *Eichhornia crassipes*. Oecologia 1990:84.
- [14] Herfjord T, Osthagen H, Saelthun NR. The water hyacinth. Norwegian Agency for Development Cooperation. Water Resources and Energy Administration, Oslo. ISBN 1994;82-410- 0207-6.
- [15] Sooknah RD, Wilkie AC. Nutrient removal by floating aquatic macrophytes cultured in anaerobically digested flushed dairy manure wastewater. Ecol Eng 2004;22:27–42.
- [16] Aquatic Ecosystem Restoration Foundation. Best Management Practices in Support of Fish and Wildlife Habitat. 3272, Sherman Ridge Road Marietta, Georgia; 2005. p. 30064.

- [17] Wilson JR, Rees M, Holst N, Thomas MB, Hill G. Water hyacinth population dynamics. Biological and integrated control of water hyacinth, *Eichhornia* crassipes. ACIAR Proc 2001;102:96–104.
- [18] Olivares E, Colonnello G. Salinity gradient in the Manamo river, a dammed distributary of the Orinoco Delta, and its influence on the presence of Eichhornia crassipes and Paspalum repens. Interciencia 2000;25:242–8.
- [19] Jafari N. Ecological and socio-economic utilization of water hyacinth (Eichhornia crassipes Mart Solms). J Appl Sci Environ Manag 2010;14(2): 43–49.
- [20] De Casabianca M, Laugier T, Posada F. Petroliferous wastewaters treatment with water hyacinths (Raffinerie de Provence, France): experimental statement. Waste Manag 1995;15(8):651–5.
- [21] Malik A. Environmental challenge vis a vis opportunity: the case of water hyacinth. Environ Int 2007;33:122–38.
- [22] Basha SA, Gopal KR, et al. A review on biodiesel production, combustion, emissions and performance. Renew Sustain Energy Rev 2009;13(6):1628–34.
- [23] Nigam PS, Singh A. Production of liquid biofuels from renewable resources. Prog Energy Combust Science 2011;37(1):52–68.
- [24] Nasir NF, Daud WRW, Kamarudin SK, Yaakob Z. Process system engineering in biodiesel production: a review. Renew Sustain Energy Rev 2013;22:631–9.
- [25] Fanchi JR. Energy technology and directions for the future. Boston: Elsevier; 2004; 213 (Chapter book).
- [26] OECD-FAO. Agricultural outlook, chapter 3 biofuels; 2012. p. 87–117; Bhattacharya A, Kumar P. Water hyacinth as a potential biofuel crop. EJEAFChe 2010;9(1):112–22.
- [27] Nigam JN. Bioconversion of waterhyacinth (*Eichornia crassipes*) hemicellulose acid hydrolysate to motor fuel ethanol by xylosefermenting yeast. J Biotechnol 2002;97:107–16.
- [28] Hossain R, Chowdhury MK, Yeasmin S, Mozammel MH. Production of ethanol using yeast isolates on water hyacinth and azolla. Bangladesh J Microbiol 2010;27:56–60.
- [29] LichtsFO, Industry Statistics. World fuel ethanol production. Renewable Fuels Association; 2010. Available at: http://www.ethanolrfa.org/news/entry/global-ethanol-production-to-reach-88.7-billion-litres-in-2011/ [retrieved 30.04.11].
- [30] Jinlin X, Grift TE, Hansena AC. Effect of biodiesel on engine performances andemissions. Renew Sustain Energy Rev 2011;15:1098–116.
- [31] OECD-FAO Agricultural Outlook and OECD/FAO2011/(http://www.agri-outlook.org/dataoecd/23/56/48178823.pdf\$); 2011–2020.
- [32] Yacob S, Hassan M, Shirai Y, Wakisaka M, Subash S. Baseline study of methane emission from anaerobic ponds of palm oil mill effluent treatment. Sci Total Environ 2006;366(1):187–96.
- [33] Saidur R, Abdelaziz EA, Demirbas A, Hossain MS, Mekhilef S. Areviewon biomass as a fuel for boilers. Renew Sustain Energy Rev 2011;15(5):2262–89.
- [34] Seyed Ehsan Hosseini, Wahi Mazlan A, Saber Salehirad, Seis Mohsin M. Evaluation ofpalmoil combustion characteristics by using the chemical equilibrium with application (CEA) software. Appl Mech Mater 2013;388:268–72.
- [35] Ighodalo OA, Zoukumor K, Egbon C, Okoh S, Odu K. Processing water hyacinth into biomass Briquettes for cooking purposes. J Emerg Trends Eng Appl Sci (JETEAS) 2011;2(2):305–7.
- [36] Forhadlbne MDA, Khan MZH, Sarkar MAR, Ali SM. Development of biogas processing from cow dung, poultry waste, and water hyacinth. Int J Nat Appl Sci 2013:2(1):13-7.
- [37] Palanisamy E, Manoharan N. Performance studies on vegetable oils and their derivative as alternate fuels forcompression ignition engines: an overview. In: Proceedings of the 19th national conference on IC engine and combustion; 2005. p. 95–100.
- [38] Banapurmath NR, Tewari PG, Hosmath RS. Performance and emission characteristics of a DI compression ignition engine operated on Honge, latropha and sesame oil methylesters. Renew Energy 2007;33:1982–8.
- [39] Rushang JM, Michael JP. Flow properties of bio diesel fuel blends at low temperatures. Fuel 2007;86:143–51.
- [40] Sharma YC, Singh B. Development of bio diesel from Karanja, a tree found in rural India. Fuel 2007;87:1740–2.
- [41] Venkatachalam P, Chitra P. Pilot plant for bio diesel production. Hand book for bio diesel. Tamil Nadu: Agricultural University; 2007; 122–38.
- [42] Chuang YS, Lay CH, Sen B, Chen CC, Gopalakrishnan K, Wu JH, et al. Biohydrogen and biomethane from water hyacinth (*Eichhornia crassipes*) fermentation: effects of substrate concentration and incubation temperature. Int J Hydrogen Energy 2011;36:14195–203.
- [43] Bolenz S, Omran H, Gierschner K. Treatment of water hyacinth tissue to obtain useful products. Bio wastes 1990;33:263–74.
- [44] Gutiérrez LE, Huerto RD, Saldaña FP, Arreguin F. Strategies for waterhyacinth (*Eichhornia crassipes*) control in Mexico. Hidrobiologia 1996;340:118–85.
- [45] Gressel J. Transgenics are imperative for biofuel crops. Plant Sci 2008;174: 246–63.
- [46] Poddar K, Mandal L, Banerjee GC. Studies on water hyacinth (Eichhornia crassipes) chemical composition of the plant and water from different habitats. Ind Vet J 1991;68:833–7.
- [47] Ikhtyar O, Shmsuzzamana F. Kinetics of gas formation from water hyacinth biomass. In: Proceedings of the international conference on water hyacinth, Hyderabad, India; 1984. p. 475–85.
- [48] Dhahiyat Y. Siregar, H. Indriati and F. Salem. Studies on the uses of water hyacinth as biogas energy resource in the dam of curag (west java). In: Proceedings of the international conference on water hyacinth, Hyderabad, India; 1984. p. 604–24.

- [49] Masami GO, Usui I, Urano N. Ethanol production from the water hyacinth Eichhornia crassipes by yeast isolated from various hydrospheres. Afr J Microbiol Res 2008;2:110–3.
- [50] Sandip S, Sandeep M, More M, Nadaf AA. Biochemical conversion of acidpretreated water hyacinth (*Eichhornia crassipes*) to alcohol using PichiaStipitis NCIM3497. Int J Adv Biotechnol Res 2012;3(2):585–90.
- [51] Clark TA, Mackie KL. Fermentation inhibitors in wood hydrolysates derived from wood *Pinus radiata*. J Chem Technol Biotechnol 1984;34b:101–10.
- [52] Sarker H, Haque KH, Alam AK, Slam R. Production of biogas from water hyacinth. In: Proceedings of the international conference on water hyacinth, India; 1984. p. 489–92.
- [53] Ando S, Arai F, Kiyoto K, Hanai S. Identification of aromatic monomers in steam-exploded polar and their influence on ethanol fermentation by Saccharomyces cereisiae. J Ferment Technol 1986;64:567–76.
- [54] Beck MJ. Factors affecting efficiency of biomass fermentation to ethanol. Biotechnol Bioeng Symp 1986;17:617–27.
- [55] Tran AV, Chambers RP. Ethanol fermentation of red oak acid prehydrolysate by the yeast *Pichia stipitis* CBS 5776. Enzyme Microb Technol 1986;8:439–44.
- [56] Nishikawa N, Sutcliffe R, Saddler JN. The influence of lignin degradation products on xylose fermentation by Klebsiella pneumoniae. Appl Microbiol Biotechnol 1988;27:549–52.
- [57] Van ZC, Prior BA, Preez JC. Acetic acid inhibition of p-xylose fermentation by Pichia stipitis. Enzyme Microb Technol 1991;13:82–6.
- [58] Dominguez H, Nunez MJ, Chamy R, Lema J. Determination of kinetic parameters of fermentation processes by a continuous unsteady-state method: application to the alcoholic fermentation of p-xylose by Pichia stipitis. Biotechnol Bioeng 1993;41:1129–32.
- [59] Aswathy US, Sukumaran RK, Lalitha DG, Rajasree KP, Singhania RR, Pandey A. Bio-ethanol from water hyacinth biomass: an evaluation of enzymatic saccharification strategy. Bioresour Technol 2010;101:925–30.
- [60] Ali N, Chaudhary BL, Khandelwal SK. Better use of water hyacinth for fuel, manure and pollution free environment. Indian J Environ Prot 2004;24: 297–303
- [61] Kumar S. Studies on efficiencies of bio-gas production in anaerobic digesters using water hyacinth and night-soil alone as well as in combination. Asian J Chem 2005;17:934–8.
- [62] Kumar A, Singh LK, Sanjoy G. Bioconversion of lignocellulosic fraction of water-hyacinth (*Eichhornia crassipes*) hemicellulose acid hydrolysate to ethanol by *Pichia stipitis*. Bioresour Technol 2009;100:3293–7.
- [63] Ma Fuying, Yang Na, Xu Chunyan, Yu Hongbo, Wu Jianguo, Xiaoyu Z. Combination of biological pretreatment with mild acid pretreatment for enzymatic hydrolysis and ethanol production from water hyacinth. Bioresour Technol 2010: 101-9600-4
- [64] Isarankura-Na-Ayudhya C, Tantimongcolwat T, Kongpanpee T, Prabkate P, Prachayasittikul V. Appropriate technology for the bioconversion of water hyacinth (*Eichhornia crassipes*) to liquid ethanol: future prospects for community strengthening and sustainable development. EXCLI | 2007;6:167–76.
- [65] Sornvoraweat B, Kongkiattikajorn J. Separated hydrolysis and fermentation of water hyacinth leaves for ethanol production. KKU Res J 2010;15:9.
- [66] Eshtiaghi MN, Yoswathana N, Kuldiloke J, Ebadi AG. Preliminary study for bioconversion of water hyacinth (*Eichhornia crassipes*) to bioethanol. Afr J Biotechnol 2012;11(21):4921–8.
- [67] Idrees M, Adnan A, Sheikh S, Qureshi FA. Optimization of dilute acid pretreatment of water hyacinth biomass for enzymatic hydrolysis and ethanol production. EXCLI J 2013;12:30–40.
- [68] Grover PD, Mishra SK. Biomass briquetting: technology and practices. Regional wood energy Development Programme in Asia, GCP/RAS/154/ NET. Bangkok: Food and Agricultural Organization of the United Nations; 1996.
- [69] Nasrin AB, Ma AN, Choo YM, Mohamad S, Rohaya MH, Azali A, et al. Oil palm biomass as potential substitution raw materials for commercial biomass briquettes production. Am J Appl Sci 2008;5(3):179–83.
- [70] Ndimele PE. The effects of water hyacinth (Eichhornia crassipes [Mart.] Solms) infestation on the physico-chemistry, nutrient and heavy metal content of Badagry Creek and Ologe Lagoon, Lagos, Nigeria. J Environ Sci Technol 2012;5(2):128–36.
- [71] Abakr YA, Abasaeed AE. Experimental evaluation of a conical-screw briquetting machine for the briquetting of carbonized cotton stalks in Sudan. J Eng Sci Technol 2006;1(2):212–20.
- [72] Kirzan P, Soos L, Matus M, Svatek M, Vukelic D. Evaluation of measured data from research of parameters impact on final briquettes density. J Appl Math 2010;3(3):70–6.
- [73] Wamukonya L, Jenkins B. Durability and relaxation of sawdust and wheat straw briquettes as possible fuels for Kenya. Biomass Bioenergy 1995; 8(3):175–9.
- [74] Frank Dr, Oroka O, Thelma AE. Fuel briquettes from water hyacinth-cow dung mixture as alternative energy for domestic and agro-industrial applications. J Energy Technol Policy 2013;3(6):56-63.
- [75] Mann MK, Spath PL. A life cycle assessment of biomass cofiring in a coalfired power plant. Clean Prod Proc 2001;3:81–91.
- [76] Narayanan KV, Natarajan E. Experimental studies on cofiringof coal and biomass blends in India. Renew Energy 2007;32(15):2461–642.
- [77] Demirbas A. Biomass resource facilities and biomass conversion processing for fuels and chemicals. Energy Convers Manag 2001;42(11):1357–78.
- [78] Hughes E. Biomass cofiring: economics, policy and opportunities. Biomass Bioenergy 2000;19(6):457–65.

- [79] Felfli FF. Biomass briquetting and itsperspectives in Brazil. Biomass Bio energy 2011;35:236–42.
- [80] Hendra D. The Utilization of water hyancinth (*Eichornia crassipes*) for basic comodity of briquette in order to alternative fuel. Darmaga Bogor: Fakultas Perikanan dan Ilmu Kelautan. Institut Pertanian Bogor II; 2011; 189–204.
- [81] Sotannde OA, Oluyege AO, Aba GB. Physical and combustion of charcoal briquettes from neem wood residues. Int Agrophys 2010;24:189–94.
- [82] Olorunnisola AO. Production of fuel briquettes from waste paper and coconut husk admixtures. Agric Eng Int: CIGR Ej 2007;1 (Manuscript EE 06 066). Davies Abolude; JSRR, Article No. JSRR.2013.009120.
- [83] Bhattacharya SC, Bhatia R, Islam MN, Shah N. Densified biomass in Thailand: potential, status and problems. Biomass 1998;8:255–66.
- [84] Kuti OA. Performance of composite sawdust briquette fuel in a biomass stove under simulated condition. AU J T 2009;12:284–8.
- [85] Kuti OA. Impact of charred palm kernel shell on the calorific value of composite sawdust briquette. J Eng Appl Sci 2007;2:62–5.
- [86] Davies RM, Abolude DS. Ignition and burning rate of water hyacinth briquettes. J Sci Res Rep 2012;2(1):111–20.
- [87] Chen L, Xing L, Lujia H. Renewable energy from agro-residues in China: solid biofuels and biomass briquetting technology. Renew Sustain Energy Rev 2009;13:2689–95.
- [88] Purohit P, Tripathi AK, Kandpal TC. Energetics of coal substitution by briquettes of agricultural residues. Energy 2006;31:1321–31.
- [89] Rotimi MD, Davies OA, Mohammed US. Combustion characteristics of traditional energy sources and water hyacinth briquettes. Int J Sci Res Environ Sci (IJSRES) 2013;1(7):144–51.
- [90] Emerhi EA. Physical and combustion properties of briquettes produced from sawdust of three hardwood species and different organic binders. Adv Appl Sci Res 2011;2(6):236–46.
- [91] Tran TT, Nguyen VD, Do DN, Nguyen HP, Choi J. Assessment of electric power generation via water hyacinths and agricultural waste. J Energy Power Eng 2011;5:627–31.
- [92] Supatata N, Buates J, Hariyanont P. Characterization of fuel briquettes made from sewage sludge mixed with water hyacinth and sewage sludge mixed with sedge. Int J Environ Sci Dev 2013;4(2).
- [93] Rodrigues AJ, Odero MO, Hayombe PO, Akuno W, Kerich D, Maobe I. Converting water hyacinth to briquettes: a beach community based approach. Int J Sci: Basic Appl Res (IJSBAR) 2014;15(1):358–78.
- [94] Dixit A, Dixit S, Goswami CS. Process and plants for wastewater remediation: a review. Sci Rev Chem Commun 2011;11:71–7.
- [95] Mahamadi C, Nharingo T. Competitive adsorption of Pb2+, Cd2+ and Zn2+ ions onto *Eichhornia crassipes* in binary and ternary systems. Bioresour Technol 2010:101:859–64.
- [96] Rahman M, Hasegawa H. Aquatic arsenic: phytoremediation using floating macrophytes. Chemosphere 2011;83:633–46.
- [97] Smolyakov B. Uptake of Zn, Cu, and Cd by water hyacinth in the initial stage of water system remediation. Appl Geochem 2012;27(6):1214–9. http://dx.doi.org/10.1016/j.apgeochem.2012.02.027.
- [98] Valero MAC, Johnson M, Mara DD. Enhanced phosphorus removal in a waste stabilization pond system with blast furnace slag filters. In: Proceedings of the second international conference smallwat. Seville. Spain: 2007.
- [99] Du Preez, Bosch JC, Prior M, Oso BA. The fermentation of hexose and pentose sugars by *Candida shehatae* and *Pichia stipitis*. Appl Microbiol Biotechnol 1986;23:228–33.
- [100] Mergaert K, Vanderhaegen B, Verstraete W. Applicability and trends of anaerobic pre-treatment of municipal wastewater. Water Res 1992;26(8): 1025–33.
- [101] Seghezzo L, Zeeman G, Van Lier JB, Hamelers HVM, Lettinga G. A review: the anaerobic treatment of sewage in UASB and EGSB reactors. Bioresour Technol 1998:65:175–90
- [102] Kiran M, Srivastava J, Tripathi B. Capacidad de retiro del nitrógeno y el fósforo de cuatro plantas elegidas en las charcas de aguas dulces tropicales. D Conserv Ambient 1991:18:143-7.
- [103] Oso B.A. Invasion of Nigeria waterways by water hyacinth: ecological and biological observation. In: Proceedings of the international workshop/seminar on water hyacinth, Lagos; 1988. p. 116–23.
- [104] Williams SR. Apparatus and method for agricultural animal wastewater treatment. United States Patent Application Number 2009. p. 250–393.
- [105] USEPA. Design manual constructed wetlands and aquatic systems for municipall wastewater treatment. U.S. Environmental Protection Agency. Report No. EPA/625/1-88/022. Office of Research and Development, Cincinnati, OH, 83, 1988.
- [106] Yang DQ, Jing YX, Chen ZP, Cheng HQ. Study on the removal effect and regulation of *Cleistocalyx operculatus* to N and P in a eutrophic water body. Acta Sci Circumst 2001;21(5):637–9.
- [107] Vaillant N, Monnet F, Sallanon H, Coudret C, Hitmi A. Treatment of domestic wastewater by an hydroponic NFT system. Chemosphere 2003;50(1):121–9.
- [108] Busk TA, Peterson JE, Reddy KR. Use of aquatic and terrestrial plants for removing phosphorus from dairy wastewaters. Ecol Eng 1995;52(3) 371–90
- [109] Costa RHR, Bavaresco ASL, Medri W, Philippi LS. Tertiary treatment of piggery wastes in water hyacinth ponds. Water Sci Technol 2000;42(10):211–4.
- [110] Wu D, Wang ZF, Feng L. The harm of over-reproduction of water hyacinth and preventative measures. Environ Sci Technol 2001;24:35–7.
- [111] Aoyama I, Nishizaki H. Uptake of nitrogen and phosphate and water purification by water hyacinth. Water Sci Technol 1991;28(7):47–53.

- [112] Jebanesan A. Bilological treatment of dairy waste by Eichhornia crassipes Solms. Environ Ecol 1997;15(3):521–3.
- [113] Jayaweera MW, Kasturiarachchi JC, Kularatne RK, Wijeyekoon SL. Contribution of water hyacinth (*Eichhornia crassipes* (Mart. Solms) grown under different nutrient conditions to Fe-removal mechanisms in constructed wetlands. J Environ Manag 2008;87(3):450–60.
- [114] Sinha AK, Sinha RK. Sewage management by aquatic weeds (water hyacinth and duckweed): economically viable and ecologically sustainable biomechanical technology. Environ Educ Inf 2000;19:215–26.
- [115] Tripathy BD, Upadhyay AR. Dairy effluent polishing by aquatic macrophytes. Water Air Soil Pollut 2003;9(143):377–85.
- [116] Jianbo Lu, Zhihui Fu, Zhaozheng Yin. Performance of a water hyacinth (Eichhornia crassipes) system in the treatment of wastewater from a duck farm and the effects of using water hyacinth as duck feed. J Environ Sci 2008:20:513-9.
- [117] Chen X, Xi Chen, Xi Wan, Weng B, Huang Q. Water hyacinth (Eichhornia crassipes) waste as an adsorbent for phosphorus removal from swine wastewater. Bioresour Technol 2010;101:9025–30.
- [118] Akinbile CO, Yusoff MS. Assessing water hyacinth (*Eichhornia crassopes*) and lettuce (*Pistia stratiotes*) effectiveness in aquaculture waste water treatment. Int J Phytoremediat 2012;14:201–11.
- [119] Rugh CL, Wilde HD, Stack NM, Thompson DM, Summers AO, Meagher RB. Mercuric ion reduction and resistance in transgenic Arabidopsis thaliana plants expressing a modified bacterial merAgene. Proc Natl Acad Sci USA 1996;93(8):3182–8187.
- [120] Srivastava AK, Purnima X. Phytoremediation for heavy metals-a land plant based sustainable strategy for environmental decontamination. Proc Natl Acad Sci India Sect B Biol Sci 1998;68(3–5):199–215.
- [121] Lasat MM. Phytoextraction of toxic metals: a review of biological mechanisms. J Environ Qual 2002;31(1):109–20.
- [122] Hinchman RR, Cristina NM, Gatliff EG. Phytoremediation: using green plants to clean up contaminated soil, groundwater, and wastewater. Illinois: Argonne National Laboratory; 1996; 1–13.
- [123] Raskin I, Smith RD, Salt DE. Phytoremediation of metals: using plants to remove pollutants from the environment. Curr Opin Biotechnol 1997;8: 221–6
- [124] Kelley C, Mielke R, Dimaquibo D, Curtis A, Dewitt J. Adsorption of Eu(III) onto roots of water hyacinth. Environ Sci Technol 1999;33(9):1439–43.
- [125] Delgado M, Bigeriego M, Guardiola E. Uptake of Zn, Cr and Cd by water hyacinths. Water Res 1993;27(2):269–72.
- [126] Klumpp A, Bauer K, Franz-Gerstein C, de Menezes M. Variation of nutrient and metal concentrations in aquatic macrophytes along the Rio Cachoeira in Bahia (Brazil), Environ Int 2002:28:165–71.
- [127] Misbahuddin M, Fariduddin A. Water hyacinth removes arsenic from arseniccontaminated drinkingwater. Arch Environ Health 2002;57(6):516–8.
- [128] Shaban W, Rmalli A, Harrington CF, Ayub M, Haris PI. Abiomaterial based approach for arsenic removal from water. J Environ Monit 2005;7:279–82.
- [129] Chigbo FE, Smith RW, Shore FL. Uptake of arsenic, cadmium, lead and mercury from polluted waters by the water hyacinth *Eichornia crassipes*. Environ Pollut Ser A Ecol Biol 1982;27(1):31–6.
- [130] Low KS, Lee CK. Removal of arsenic from solution by water hyacinth (*Eichhornia crassipes* (Mart Solms). Pertanika 1990;13(1):129–31.
- [131] Ingole NW, Bhole AG. Removal of heavy metals from aqueous solution by waterhyacinth (*Eichhornia crassipes*). J Water SRT – Aqua 2003;52:119–28.
- [132] Liao S, Chang W. Heavy metal phytoremediation by water hyacinth at constructed wetlands in Taiwan. J Aquat Plant Manag 2004;42:60–8.
- [133] Xiaomei L, Kruatrachue M, Pokethitiyook P, Homyok K. Removal of cadmium and zinc by water hyacinth, *Eichhornia crassipes*. ScienceAsia 2004;30: 93–103
- [134] Mahmood Q, Zheng P, Islam E, Hayat Y, Hassan MJ, Jilani G, et al. Lab scale studies on water hyacinth (*Eichhornia crassipes* Marts Solms) for biotreatment of textile wastewater. , Casp J Env Sci 2005;3(2):83–8.
- [135] Anzeze AD. Biosorption of heavy metals using water hyacinth *Eichhornia crassipes* (Mart.) solms-Laubach: adsorption properties and technological assessment. Online available at: (http://erepository.uonbi.ac.ke:8080/xmlui/handle/123456789/6844); 2011.
- [136] Sannigrahi AK, Chakrabortty S, Borah BC. Large scale utilization of water hyacinth (Eichhornia crassipes) as raw material for vermi composting and surface mulching in vegetable cultivation. Ecol Environ Conserv 2002;8: 269–71
- [137] Stocker RK, Haller WT. Residual effects of herbicide-treated Eichhornia crassipes used as a soil amendment. Hydrobiologia 1999;415:329–33.
- [138] Akcin G, Saltabas O, Afsar H. Removal of lead by water hyacinth (*Eichhornia crassipes*). J. Environ. Sci. Health Part A 1994;A29(10):2177–83.
- [139] Lytle C, Lytle F, Yang N, Qian J, Hansen D, Zayed A. Reduction of Cr(VI) to Cr (III) by wetland plants: potential for in situ heavy metal detoxification. Environ Sci Technol 1998;32(20):3087–93.
- [140] Lenzi E, Luchese E, de Lima Barbosa S. Improvement of the *Eichhornia crassipes* water hyacinth-use in the decontamination of chromium contaminated solution. Arq Biol Technol 1994;37(3):603–9.
- [141] Woomer PL, Muzira R, Bwamiki D, Mutetikka D, Amoding A, Bekunda MA. Biological management of water hyacinth waste in Uganda. Biol Agric Hortic 2000;17:181–96.
- [142] Gunnarsson CC, Petersen CM. Water hyacinths as a resource in agriculture and energy production: a literature review. Waste Manag 2007;27(1): 117–29.

- [143] Oguniade Y, AfolabiO A, Osuntogun OA, OkeO L. Extraction of protein fromwater hyacinth. In: Proceedings of the internationalworkshop/seminar on water hyacinth, Lagos, 7–12 August 1988. p. 1.
- [144] Igbinosun JE, Talabi SO. Studies on the nutrition of brackish water catfish: C'hrysichthys nigrodigitaius. In: Paper presented at the 2nd conference of fisheries society of Nigeria, Calabar; 1982.
- [145] Abbasi SA, Ramasamy EV. Utilization of biowaste solids by extracting volatile fatty acids with subsequent conversion to methane and manure. In: Proceedings of the 12th international conference on solid waste technology and management, Philadelphia; 1996. p. 4C1–C8.
- [146] Lu J, Zhu L, Hu G, Wu J. Integrating animal manure-based bioenergy production with invasive species control: a case study at Tongren pig farm in China. Biomass Bioenergy 2010;34:821–7. http://dx.doi.org/10.1016/j.biombioe.2010.01.026.
- [147] Saha S, Ray Ak. Evaluation of nutritive value of water hyacinth (*Eichhornia crassipes*) Leaf Meal in compound diets for Rohu, Labeo rohita (Hamilton, 1822) fingerlings after fermentation with two bacterial strains isolated from fish gut. Turk J Fish Aquat Sci 2011;11:199–207. http://dx.doi.org/10.4194/trjfas.2011.0204.
- [148] Dai YL. Observation and improvement of general egg quality. Chin J Poult 2001;23(10):42–3.
- [149] Van Der Meer MB, Verdegem MCJ. Comparison of amino acid profiles of feeds and fish as a quick method for selection of feed ingredients: a case study of *Colossoma macropomum* (Cuvier). Aquac Res 1996;27:487–95.
- [150] Kivaisi AK, Mtila M. Chemical composition and in vitro degradability of whole plants and shoots of the water hyacinth (*Eicchornia crassipes*) by rumen micro-organisms. Tanzan Vet J 1995;15:121–9.